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- 1 Leong CC, Marley JV, Loh S, Milech N, Robinson BW, Garlepp MJ. Transfection of the gene for B7-1 but not B7-2 can induce immunity to murine malignant mesothelioma. *Int J Cancer* 1997; **71**: 476–82.
- 2 Alley EW, Lopez J, Santoro A, et al. Clinical safety and activity of pembrolizumab in patients with malignant pleural mesothelioma (KEYNOTE-028): preliminary results from a non-randomised, open-label, phase 1b trial. *Lancet Oncol* 2017; **18**: 623–30.
- 3 Hassan R, Thomas A, Patel MR, et al. Avelumab (MSB0010718C; anti-PD-L1) in patients with advanced unresectable mesothelioma from the JAVELIN solid tumor phase 1b trial: safety, clinical activity, and PD-L1 expression. *Proc Am Soc Clin Oncol* 2016; **34** (suppl 15): 8503.
- 4 Quispel-Jansen J, Zago G, Schouten R, et al. A phase II study of nivolumab in malignant pleural mesothelioma (NivoMes): with translational research (TR) biopsies. World Conference on Lung Cancer; Vienna; Dec 4–7, 2016. OA13.01.
- 5 Kindler HL. Phase II trial of pembrolizumab in patients with malignant mesothelioma (MM): interim analysis. World Conference on Lung Cancer; Vienna; Dec 4–7, 2016. OA13.02.
- 6 Scherpereel A, Mazieres J, Greillier L, et al. Second- or third-line nivolumab (Nivo) versus nivo plus ipilimumab (ipi) in malignant pleural mesothelioma (MPM) patients: results of the IFCT-1501 MAPS2 randomized phase II trial. *Proc Am Soc Clin Oncol* 2017; **35** (suppl 18): LBA8507.
- 7 Scherpereel A, Wallyn F, Albelda SM, Munck C. Novel therapies for malignant pleural mesothelioma. *Lancet Oncol* 2018; **19**: e161–72.
- 8 Calabrò L, Morra A, Giannarelli D, et al. Tremelimumab combined with durvalumab in patients with mesothelioma (NIBIT-MESO-1): an open-label, non-randomised, phase 2 study. *Lancet Respir Med* 2018; **6**: 451–60.
- 9 Wolchok JD, Chiarion-Sileni V, Gonzalez R, et al. Overall survival with combined nivolumab and ipilimumab in advanced melanoma. *N Engl J Med* 2017; **377**: 1345–56.



Cough swabs less useful but induced sputum very useful in symptomatic older children with cystic fibrosis



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Microbiological surveillance continues to be an important aspect of cystic fibrosis care.^{1,2} In young children, such surveillance is complicated by the inability to expectorate sputum. Thus, cough swabs are routinely used to obtain samples, and some cystic fibrosis centres use regular bronchoalveolar lavage or induced sputum sampling.

In *The Lancet Respiratory Medicine*, Katherine Ronchetti and colleagues³ compared the diagnostic yield of bacterial culture results from 167 paired induced sputum and cough swab samples in patients aged 6 months to 18 years.³ In a subset of their study they compared bacterial yield from induced sputum to bronchoalveolar lavage. Induced sputum samples had a significantly higher bacterial yield than cough swab samples: 86 different pathogens were isolated in total from the samples, and 79 [92%] of these were identified by sputum induction whereas 27 [31%] were on cough swab samples ($p < 0.0001$). This difference suggests that induced sputum can detect pathogens from the lower airways that cough swabs cannot. A strength of this first stage of the study was high participant numbers: 124 patients were recruited from whom 167 paired samples were successfully taken. Children younger than 6 years were also well represented in this part of the study ($n = 72$). In view of these results, induced sputum appears to be far superior to cough swab for detection of lower airway pathogens in patients with cystic fibrosis. These findings are useful and consistent

with other studies that showed improved microbial yield of induced sputum over upper-airway sampling in children.^{4,5} The poor diagnostic value of cough swabs is also consistent with other findings that oropharyngeal swabs bear little-to-no relation to lower-airway pathology in young children with cystic fibrosis.⁶

In a subset of largely symptomatic patients, Ronchetti and colleagues compared bacterial culture yield from 41 paired induced sputum samples and single-lobe, two-lobe, and six-lobe bronchoalveolar lavages. Sequentially more pathogens were isolated with bronchoalveolar lavage as more lobes were lavaged. Sputum induction isolated 27 (69%) of the 39 total pathogens detected by both techniques versus 22 (56%) on single-lobe bronchoalveolar lavage, 28 (72%) on two-lobe bronchoalveolar lavage, and 33 (85%) on six-lobe bronchoalveolar lavage. Because different pathogens were detected between induced sputum and bronchoalveolar lavage, Ronchetti and colleagues³ advocate for a combination of induced sputum and six-lobe lavage as the new standard of care for lower airway pathogen surveillance. They also make a case for the use of induced sputum as a non-invasive surrogate for bronchoalveolar lavage in symptomatic patients and propose that bronchoalveolar lavage should be reserved for patients with persistent signs or symptoms not explained by sputum induction. In symptomatic patients, induced sputum will correctly describe pathogens in almost two-thirds of cases

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and, if correctly used, might reduce the need for bronchoscopy.

Importantly, five (13%) of the total 39 pathogens identified were only detected by the induced sputum technique. This highlights an issue often encountered in diagnostic studies, of how to best define the gold standard. For example, bacteria found only from cough swabs, but not lower airway samples, will easily be discounted as normal commensal throat flora. The same suggestion could be made for induced sputum, as these samples pass the upper airways during collection. However, the authors suggest that the pathogens that were identified solely from induced sputum might be from the large intrathoracic airways, which are bypassed in a bronchoalveolar lavage procedure. This seems reasonable, particularly as bacterial bronchitis is common in symptomatic children with cystic fibrosis. Rather than discarding the pathogens that were exclusively found on induced sputum, the authors have included them in a combined gold standard, thereby highlighting the additional value that sputum induction might have for pathogen detection in symptomatic children with cystic fibrosis.

Care should be taken to not simply extrapolate results from the smaller subgroup of Ronchetti and colleagues' study.³ Only small numbers of children younger than 6 years were recruited, and the sample was too small to enable a separate analysis of children younger than 6 years. Also, for most samples taken in the part of the study comparing sputum induction with bronchoalveolar lavage, participants were symptomatic (ie, had respiratory exacerbations).

The distinction between asymptomatic young children and symptomatic older children is important. Cystic fibrosis lung disease starts in the small airways and in young children, infection might be present in the absence of symptoms.⁷ Induced sputum might miss infection that will lead to structural lung damage in young asymptomatic children. Indeed, the largest study thus far that compared 61 paired sputum

induction and bronchoalveolar lavage samples from young asymptomatic children suggested that induced sputum samples only had a 36.8% (95% CI 16.3–61.6) sensitivity to detect microbiota from the lower airway in cystic fibrosis using bronchoalveolar lavage as the gold-standard method.⁸

Infancy and the early preschool years comprise a vital period when structural changes in the lungs of people with cystic fibrosis begins.⁷ Therefore, while Ronchetti and colleagues³ provide robust evidence for the usefulness of sputum induction in symptomatic older children with cystic fibrosis, future efforts should be focused on developing an accurate non-invasive method for microbiological surveillance in young asymptomatic children.

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- 1 Smyth AR, Bell SC, Bojcin S, et al. European Cystic Fibrosis Society standards of care: best practice guidelines. *J Cyst Fibros* 2014; **13** (suppl 1): S23–42.
- 2 Elborn JS. Cystic fibrosis. *Lancet* 2016; **388**: 2519–31.
- 3 Ronchetti K, Tame J-D, Paisey C, et al. The CF-Sputum Induction Trial (CF-SpIT) 1 to assess lower airway bacterial sampling in young children with cystic fibrosis: a prospective internally controlled interventional trial. *Lancet Respir Med* 2018; **6**: 461–71.
- 4 Muloiwa R, Dube FS, Nicol MP, Zar HJ, Hussey GD. Incidence and diagnosis of pertussis in South African children hospitalized with lower respiratory tract infection. *Pediatr Infect Dis J* 2016; **35**: 611–16.
- 5 Zampoli M, Pillay K, Carrara H, Zar HJ, Morrow B. Microbiological yield from induced sputum compared to oropharyngeal swab in young children with cystic fibrosis. *J Cyst Fibros* 2016; **15**: 605–10.
- 6 Breuer O, Caudri D, Akesson L, Ranganathan S, Stick SM, Schultz A. The clinical significance of oropharyngeal cultures in young children with cystic fibrosis. *Eur Respir J* 2018; published online April 20. DOI:10.1183/13993003.00238-2018.
- 7 Ranganathan SC, Hall GL, Sly PD, Stick SM, Douglas TA. Early lung disease in infants and preschool children with cystic fibrosis. What have we learned and what should we do about it? *Am J Respir Crit Care Med* 2017; **195**: 1567–75.
- 8 D'Sylva P, Caudri D, Shaw N, et al. Induced sputum to detect lung pathogens in young children with cystic fibrosis. *Pediatr Pulmonol* 2017; **52**: 182–89.